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## ESTIMATION OF CHLOROPHYLL AND CAROTENOID CONTENTS OF WHOLE TOMATO BY LIGHT ABSORBANCE TECHNIQUE

### ABSTRACT

The relationship between optical measurements of whole tomato and pigment content was studied. The difference between the absorbances (A) at 710 and 780 nm [ $\Delta A$  (710–780 nm)] correlated significantly ( $r = 0.98$ ) with chlorophyll contents of 0.3–13.4  $\mu\text{g/g}$  fresh weight (gfw). The  $\Delta A$  (570–780 nm) correlated significantly ( $r = 0.97$ ) with lycopene contents of 0.2–46.7  $\mu\text{g/gfw}$ . The  $\Delta A$  (550–580 nm) correlated significantly with the  $\beta$ -carotene content ( $r = 0.95$ ). The reflectance measurements correlated with the chlorophyll and lycopene contents, but the coefficients were lower and the standard errors were larger than those observed with the absorbance measurements.

that cannot be sacrificed for chemical analyses of the pigments.

In this study, pigment contents were correlated with absorbances at wavelengths specified by Worthington (1974). Other wavelengths were also evaluated so that the best correlation might be found.

Additionally, since light reflectance measurements are used universally to describe tomato color, the relationships of light reflectance to absorbance and to pigment content were determined.

### INTRODUCTION

WORTHINGTON (1974) estimated the ripeness of externally green and red tomatoes on the basis of absorbances (A) at 510, 600 and 690 nm. (A =  $\log 1/T$ , where T is the transmittance of the sample. Absorbance as defined here includes the loss from absorption, reflectance and scatter of light.) Light absorption was assumed to be due to chlorophyll in green tomatoes and carotenoids in red tomatoes. If this assumption is correct, pigment content could be estimated from absorbance measurements of a whole fruit. Also, pigment content or change of content might be used as an index of physiological age of fruit

### EXPERIMENTAL

FRESHLY HARVESTED green tomatoes (cv. Walter) ranging from 5.7–6.6 cm diameter were air shipped from Miami, Fla. to Beltsville, Md. Fruit were held at 15°C in the dark until they reached the appropriate stage of ripeness for analyses. The stages were determined on the basis of criteria developed by Worthington (1974). Tomatoes in stage 1 (Table 1), termed "immature-green," lacked red internal color and gel type of placental tissue. Those in stages 2–4, termed "mature-green," had increased amounts of red internal color and gel type of placental tissue. Those in stage 5, termed "breakers," showed incipient ripening externally. Tomatoes in the remaining five stages had increased amounts of red color.

Fruit were measured optically and sorted into three lots of five fruit each for each stage of ripeness. The five fruit of similar absorbance were

Table 1—Chlorophyll and carotenoid compositions of "Walter" Tomato fruit categorized by absorbance (A) readings<sup>a</sup>

| Stages | $\Delta$ (nm) |         | Pigment content <sup>b</sup> |                   |                   |                    |                      |                   |                      |
|--------|---------------|---------|------------------------------|-------------------|-------------------|--------------------|----------------------|-------------------|----------------------|
|        | 510–600       | 600–690 | Total chlorophyll            | Lycopene          | $\beta$ -carotene | $\gamma$ -carotene | $\epsilon$ -carotene | $\zeta$ -carotene | $\beta$ -zeacarotene |
|        |               |         | $\mu\text{g/gfw}$            | $\mu\text{g/gfw}$ | $\mu\text{g/gfw}$ | $\mu\text{g/gfw}$  | $\mu\text{g/gfw}$    | $\mu\text{g/gfw}$ | $\mu\text{g/gfw}$    |
| 1      | 0.02          | +0.18   | 13.4a                        | —                 | 0.7a              | 0.03a              | 0.04a                | 0.05a             | 0.02a                |
| 2      | 0.20          | –0.30   | 11.2b                        | —                 | 0.7a              | 0.03a              | 0.04ab               | 0.05a             | 0.03a                |
| 3      | 0.61          | –0.63   | 8.6c                         | 0.2a              | 1.0ab             | 0.04a              | 0.05ab               | 0.03a             | 0.03a                |
| 4      | 1.08          | –0.93   | 5.9cd                        | 0.6a              | 1.3ab             | 0.12ab             | 0.09ab               | 0.04a             | 0.05ab               |
| 5      | 1.52          | –1.08   | 4.2d                         | 2.5a              | 1.8b              | 0.26b              | 0.07ab               | 0.05a             | 0.04ab               |
| 6      | 1.56          | –0.60   | 1.0e                         | 10.1b             | 3.1c              | 0.54c              | 0.15b                | 0.08a             | 0.06ab               |
| 7      | 1.02          | 0.22    | 0.4e                         | 16.7c             | 3.3c              | 0.87d              | 0.12ab               | 0.17ab            | 0.12bc               |
| 8      | 0.71          | 0.62    | 0.3e                         | 22.8d             | 3.7c              | 0.90d              | 0.12ab               | 0.27bc            | 0.18cd               |
| 9      | 0.32          | 1.10    | 0.3e                         | 34.8e             | 3.1c              | 0.97d              | 0.08ab               | 0.42c             | 0.21d                |
| 10     | 0.01          | 1.51    | 0.3e                         | 46.7f             | 2.9c              | 1.13e              | 0.11ab               | 0.89d             | 0.29e                |

<sup>a</sup> Average of three samples, each containing five tomatoes.

<sup>b</sup> Values within a column not followed by a common letter are significantly different ( $P < 0.01$ ) (Duncan, 1955).

analyzed together for pigment content. The average optical measurement of these fruit was compared with the pigment content.

Two instruments were used for measuring the absorbance. A four-filter photometer was used for wavelengths reported by Worthington (1974). A high-intensity spectrophotometer (Massie and Norris, 1975) was used for the other wavelengths evaluated.

#### Four-filter photometer

The four-filter photometer measures absorbance at four wavelengths on samples having absorbance as high as 9.

The absorbances of the whole tomato were measured at 510, 600 and 690 nm as described by Worthington (1974). The ripeness stages of green fruit and breaker were based on the difference between absorbances at 510 and 600 nm [ $\Delta A$  (510–600 nm)]; where as the ripeness stages of red fruit were based on  $\Delta A$  (600–690 nm) (Table 1).

#### High-intensity spectrophotometer

Tomatoes were held in the inerttube-venturi-type holder (Massie and Norris, 1975) and the absorbances were measured at 50 wavelengths spaced 10 nm apart from 390 to 880 nm. Data were stored on magnetic tape and later compared with chemical analyses of the pigments to determine the best single wavelength and best combination of two wavelengths to predict pigment content.

#### Reflectance

Light reflectance was measured with a Hunter D25 Color Difference Meter, standardized with a white standard tile. The blossom-end of the fruit was placed on a 5-cm diameter aperture. The  $L$ ,  $a_L$  and  $b_L$  values were recorded and used to calculate tristimulus  $X$ ,  $Y$  and  $Z$  (Wysecki and Stiles, 1967).

#### Pigments

Chlorophylls a and b were analyzed and calculated as described by Sweeney and Martin (1961), except that the initial maceration with acetone contained 3g  $MgCO_3$  per 150g tomato sample.

The carotenoids were analyzed as described by Tomes (1963) with a few modifications. A 50-g aliquot of the tomato acetone- $MgCO_3$  macerate that was prepared for chlorophyll analysis was homogenized with 75 ml acetone-hexane (30:45 v/v) in a Virtis homogenizer for 7 min. The chromatographic column of  $MgO$ -Hyflo Supercel was washed with 0.5% acetone in hexane to minimize isomerization of carotenoids (Wiseman et al., 1952).  $\epsilon$ -Carotene,  $\beta$ -carotene and  $\zeta$ -carotene were eluted from the column with increasing concentrations of acetone in hexane.  $\beta$ -Zeaxanthin,  $\gamma$ -carotene and lycopene were eluted from slices of the extruded column. The absorption spectrum of each eluate was measured with a Bausch & Lomb recording spectrophotometer and the concentration was calculated by using the extinction coefficient reported by Davies (1965).

## RESULTS & DISCUSSION

#### Pigments

The average chlorophyll content decreased from 13.4  $\mu\text{g/g}$  fresh weight (gfw) in immature-green (stage 1) fruit to 0.3  $\mu\text{g/gfw}$  in partially ripe (stage 8) fruit (Table 1). Changes in the chlorophyll content after stage 6 were not significant and a small amount (0.3  $\mu\text{g}$ ) of chlorophyll was still present in the table ripe (stage 10) fruit. The ratio of chlorophyll "a" to "b" was 2 in the immature fruit and approached 1 as the fruit ripened.

Carotenoid accumulation differed with time. Lycopene content at stage 3 was 0.2  $\mu\text{g/gfw}$  and increased at each successive stage to 46.7  $\mu\text{g/gfw}$  (Table 1); however, the increase from stages 3 to 5 was not significant.  $\beta$ -Carotene was 0.7  $\mu\text{g/gfw}$  at stage 1 and accumulated to a maximum of 3.7 at stage 8. Only the increase from stages 5 to 6 was significant.  $\gamma$ -Carotene was 0.03  $\mu\text{g/gfw}$  at stage 1 and increased to 1.13 by stage 10. The increases from stages 1 to 4 and from stages 7 to 9 were not significant. The quantitative increases of  $\epsilon$ -carotene,  $\zeta$ -carotene, and  $\beta$ -zeaxanthin were small, and the total level of the three was 1.29  $\mu\text{g/gfw}$  at stage 10.

#### Four-filter photometer

The measured absorbance at 510 nm changed only slightly with each successive stage of ripening. The absorbance at 690 nm decreased continuously from stages 1 to 10. The absorbance at 600 nm decreased from stages 1 to 5 and then

increased from stages 6 to 10. Thus the  $\Delta A$  (510–600 nm) increased up to stage 6 and then decreased, and the  $\Delta A$  (600–690 nm) decreased up to stage 5 and then increased (Table 1). Because of the reversal of  $\Delta A$  between stages 5 and 6, Worthington (1974) used  $\Delta A$  (510–600 nm) to measure maturity of green fruit and those showing incipient ripe color, and  $\Delta A$  (600–690 nm) for those showing ripe color.

The chlorophyll content correlated significantly with the  $\Delta A$  (510–600 nm) of fruit at stages 1 to 5 ( $r = 0.91$ ), but the coefficient was better ( $r = 0.97$ ) when the log of chlorophyll content was correlated with the  $\Delta A$  values (Fig. 1). However, the chlorophyll content of only fruit at stages 2 to 5 could be estimated satisfactorily with these absorbance values. The average chlorophyll content of fruit at stage 1 fit the regression line, but the replicate values differed widely. After stage 5 the relationship was not logarithmic.

The lycopene content correlated significantly with the  $\Delta A$  (600–690 nm) of fruits at stages 6 to 10 ( $r = 0.94$ ), but the coefficient was better ( $r = 0.98$ ) when the log of lycopene content was correlated with the  $\Delta A$  values (Fig. 2). The lycopene contents of only fruit at stages 7 to 10 could be estimated satisfactorily with  $\Delta A$  (600–690 nm). The  $\Delta A$  at stage 6 deviated from the line due to absorption at 690 nm by the chlorophyll (data not shown). The chlorophyll contents at

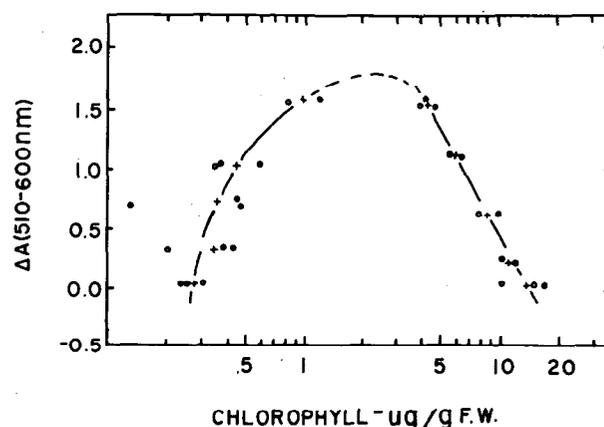


Fig. 1—Correlation between total chlorophyll content and  $\Delta A$  (510–600 nm) (four-filter photometer) of whole "Walter" tomatoes.

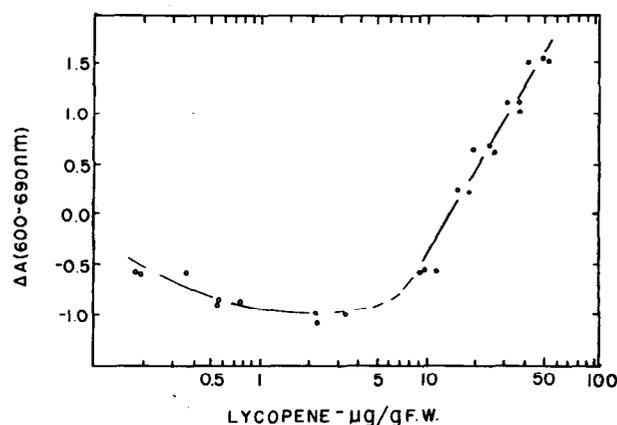


Fig. 2—Correlation between lycopene content and  $\Delta A$  (600–690 nm) (four-filter photometer) of whole "Walter" tomatoes.

stages 6 and 7 were not different statistically, but the difference did affect the absorbance.

**High-intensity spectrophotometer**

Typical spectra of fruit at different stages of ripeness (Fig. 3) show the changes in absorbance with ripening. The absorbance in the 600-710 nm region decreased during stages 1 to 6 with the loss of chlorophyll. With continued ripening of the fruit the absorbance from about 500-620 nm increased, probably due to changes in the carotenoids, particularly lycopene. Thus, from about 560-620 nm the absorbance decreased during stages 1 to 5 and then increased during stages 6 to 10. This pattern of change at 600 nm was also observed with the four-filter photometer. Absorbance values below 500 nm were not valid due to interference by filter stray light (Massie and Norris, 1975).

For the selection of wavelength pairs that correlated best with chlorophyll content, two sets of criteria were used. One was that 690 nm be used as one of the wavelengths because it was used in the four-filter photometer and by Sidwell et al. (1961) for estimating chlorophyll content. The other was that the best linear correlation be sought.

When 690 nm was one of the wavelength pair, any wavelength from 730-830 nm was a satisfactory companion, but the linear correlation was the highest with 780 nm ( $r = 0.96$ ). In using  $\Delta A (690-780 \text{ nm})$  to estimate chlorophyll content, the calculated value was higher than the actual value when the

$\Delta A$  was from 0.4-1.1 (Fig. 4). At high chlorophyll levels, the actual values deviated widely because of fluorescence (Massie and Norris, 1975). Thus 690 and 780 nm did not appear to be the best wavelength pair for estimating chlorophyll.

The  $\Delta A$  of wavelength pair, 710-780 nm, gave the highest linear correlation with the chlorophyll content (Fig. 5). The coefficient was significant (0.98), and the linear regression equation was:

$$\Delta A (710-780 \text{ nm}) = 0.0178 + 0.0389 \text{ chlorophyll}$$

The  $\Delta A (710-780 \text{ nm})$  of about -0.05 to 0.40 could be used to estimate the chlorophyll content of fruits at stages 2 through 10. The content could not be estimated satisfactorily at stage 1 because the replicates differed widely as observed with other measurements.

The lycopene content correlated linearly and significantly ( $r = 0.97$ ) with  $\Delta A (570-780 \text{ nm})$  (Fig. 6). The regression equation of the linear line was:

$$\Delta A (570-780 \text{ nm}) = 0.604 + 0.0431 \text{ lycopene}$$

The  $\Delta A (570-780 \text{ nm})$  could be used to estimate the lycopene content of fruit at any of the 10 stages in contrast to only 4 stages that were covered when  $\Delta A (600-690 \text{ nm})$  was used.

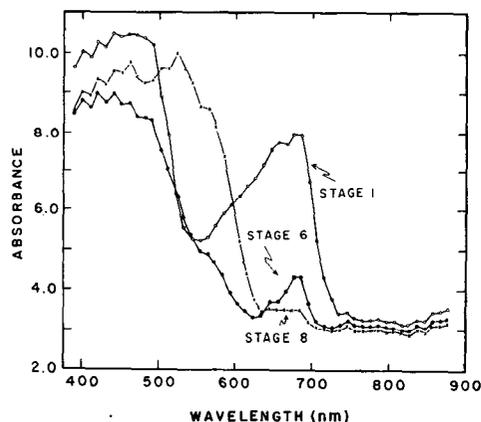


Fig. 3—Absorption spectra of green (stage 1), partially ripe (stage 6) and ripe (stage 8) tomatoes.

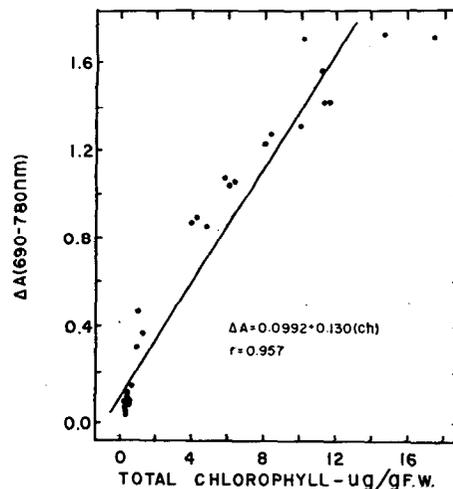


Fig. 4—Correlation between total chlorophyll content and  $\Delta A (690-780 \text{ nm})$  (high-intensity spectrophotometer) of whole "Walter" tomatoes.

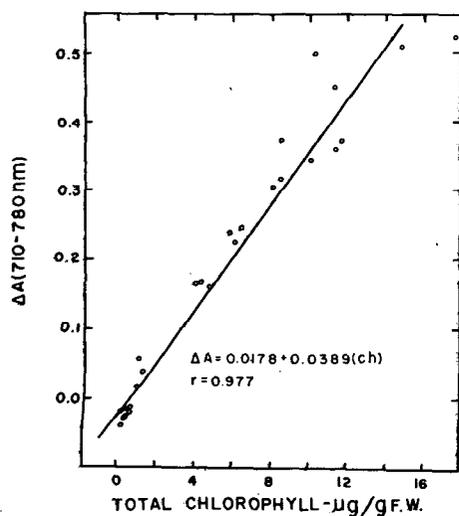


Fig. 5—Correlation between total chlorophyll content and  $\Delta A (710-780 \text{ nm})$  (high-intensity spectrophotometer) of whole "Walter" tomatoes.

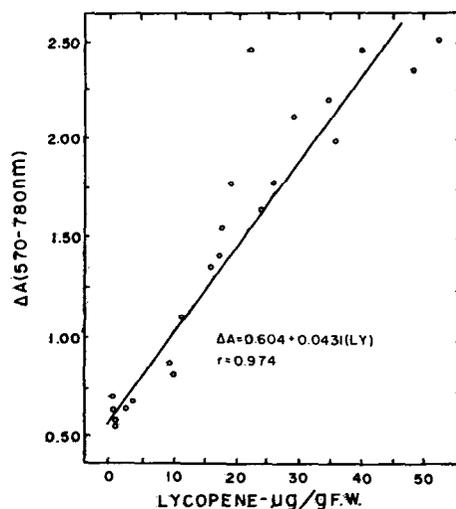


Fig. 6—Correlation between lycopene content and  $\Delta A (570-780 \text{ nm})$  (high-intensity spectrophotometer) of whole "Walter" tomatoes.

The  $\beta$ -carotene content correlated significantly [ $r = 0.95$  with  $\Delta A$  (550–580 nm)] (Fig. 7). The linear regression equation was:

$$\Delta A = -0.433 + 0.297 \beta\text{-carotene}$$

The deviation of the predicted from the actual value was large. Thus additional study is needed to reassess this relationship.

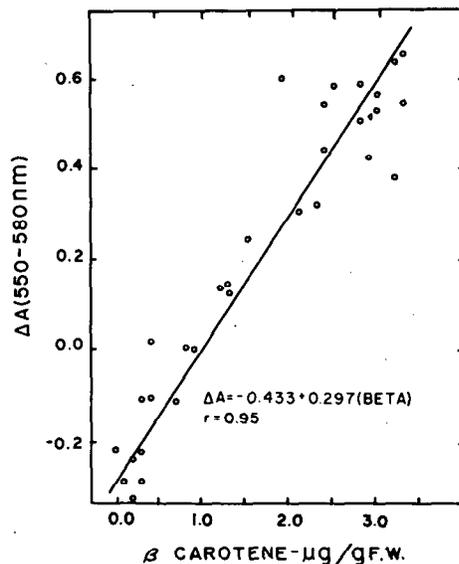


Fig. 7—Correlation between  $\beta$ -carotene content and  $\Delta A$  (550–580 nm) (high-intensity spectrophotometer) of whole "Walter" tomatoes.

Table 2—L,  $a_L$  and  $b_L$  values (Hunterlab color difference meter) of "Walter" tomatoes categorized on the basis of absorbance ( $\Delta A$ ) measurements<sup>a</sup>

| Stage | $\Delta A$ (nm) |         | Hunterlab color difference meter readings <sup>b</sup> |         |         |
|-------|-----------------|---------|--|---------|---------|
|       | 510–600         | 600–690 | L  | $a_L$   | $b_L$   |
| 1     | 0.02            | +0.18   | 55.58a   | -10.89a | 21.23c  |
| 2     | 0.20            | -0.30   | 55.80a   | -11.19a | 22.04bc |
| 3     | 0.61            | -0.63   | 56.22a   | -9.78ab | 22.94ab |
| 4     | 1.08            | -0.93   | 56.31a   | -7.57b  | 23.24a  |
| 5     | 1.52            | -1.10   | 57.00a   | -3.64c  | 23.61a  |
| 6     | 1.56            | -0.60   | 52.49b   | 9.28d   | 21.73c  |
| 7     | 1.02            | 0.22    | 47.31c   | 20.31e  | 19.49d  |
| 8     | 0.71            | 0.62    | 45.00c   | 24.90f  | 17.28e  |
| 9     | 0.32            | 1.10    | 41.34d   | 27.84g  | 15.87f  |
| 10    | 0.01            | 1.51    | 38.38e   | 30.27h  | 15.76f  |

<sup>a</sup> Average of three samples, each containing five tomatoes

<sup>b</sup> Values within a column not followed by a common letter are significantly different ( $P < 0.01$ ) (Duncan, 1955).

## Reflectance

The "L" value, which indicates lightness or darkness, increased as the green color disappeared, then decreased as the red color increased after stage 6 (Table 2). The " $a_L$ ," which indicates greenness or redness, approached zero from a negative value as the green color disappeared, then increased positive with red color formation. The " $b_L$ ," which indicates blueness or yellowness, increased up to stage 5 and then decreased.

Some of the light reflectance values correlated with the pigment content. The "L," " $a_L$ " and tristimulus "Y" and "Z" correlated significantly with the chlorophyll content. The linear correlation coefficients were 0.69, -0.85, 0.70 and 0.76, respectively. All the measurements correlated significantly with the lycopene content. The coefficients of the linear correlation were -0.96, 0.92, -0.93, -0.72, -0.95 and -0.92, respectively, for L,  $a_L$ ,  $b_L$ , X, Y and Z. Although the coefficients of L and Y approached the correlation between  $\Delta A$  (570–780 nm) and lycopene ( $r = 0.97$ ), the standard errors (SE) of L and Y (4.62 and 5.10, respectively) were larger than the SE of  $\Delta A$  (3.76). The close correlation between carotenoid content and reflectance measurements of tomatoes have been reported by others (Hall, 1963; Koskitalo and Ormrod, 1972).

## CONCLUSION

THE CHLOROPHYLL and lycopene contents in tomatoes could be estimated rapidly by nondestructive optical methods. The pigment contents in tomatoes of a limited range of maturities could be estimated when absorbance at wavelengths described by Worthington (1974) were used. A wider range of maturities was covered when  $\Delta A$  (710–780 nm) and  $\Delta A$  (570–780 nm), were used for chlorophyll and lycopene respectively. The light reflectance values could be used to estimate pigment content, but the correlation coefficients were lower and the standard errors were greater than those observed with the  $\Delta A$  values.

## REFERENCES

- Davies, B.H. 1965. Analysis of carotenoid pigments. In "Chemistry and Biochemistry of Plant Pigments," Ed. Goodwin, T.W. Academic Press, New York.
- Duncan, D.B. 1955. Multiple range and multiple F. tests. *Biometrics* 11: 1.
- Hall, C.B. 1963. Firmness and color of some tomato varieties during ripening and according to harvest dates. *Proc. Amer. Soc. Hort. Sci.* 84: 507.
- Koskitalo, L.N. and Ormrod, D.P. 1972. Effects of sub-optimal ripening temperatures on the color quality and pigment composition of tomato fruit. *J. Food Sci.* 37: 56.
- Massie, D.R. and Norris, K.H. 1975. A high-intensity spectrophotometer interfaced with a computer for food quality measurement. *Transactions ASAE* 18(1): 173.
- Sidwell, A.P., Birth, G.S., Ernest, J.V. and Golumbic, C. 1961. The use of light-transmittance techniques to estimate the chlorophyll content and stage of maturation of Elberta peaches. *Food Technol.* 15(2): 75.
- Sweeney, J.P. and Martin, M.E. 1961. Stability of chlorophyll in vegetables as affected by pH. *Food Technol.* 15(5): 263.
- Tomes, M.L. 1963. Temperature inhibition of carotene synthesis in tomato. *Bot. Gaz.* 124(3): 180.
- Wiseman, H.G., Stone, S.S., Savage, H.L. and Moore, L.A. 1952. Action of celites on carotene and lutein. *Anal. Chem.* 24: 681.
- Worthington, J.T. 1974. A light transmittance technique for determining tomato ripening rate and quality. *Acta Hort.* 1(38): 193.
- Wyszecski, G. and Stiles, W.S. 1967. "Color Science: Concepts and Methods, Quantitative Data and Formula." John Wiley and Sons, Inc., New York.
- Ms received 7/24/75; revised 9/25/75; accepted 9/30/75.

We thank Miss Judith Abbott and Mrs. Barbara Aulenbach for technical assistance.

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