

# Low O<sub>2</sub> Atmospheres Affect Storage Quality of Zucchini Squash Slices Treated with Calcium

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## ABSTRACT

Calcium chloride treated or nontreated zucchini squash slices were stored in air or low O<sub>2</sub> (0.25, 0.5 and 1%) at 10°C. Respiration rate, ethylene production, and development of browning/decay were reduced under low O<sub>2</sub>. Slices stored under 0.25% O<sub>2</sub> had less weight loss and browning/decay, and greater shear force and L-ascorbic acid content than those stored in air. Microbial count, pH, and color at the end of storage were improved by low O<sub>2</sub>. Calcium treatment had no additive effect on maintaining quality of zucchini squash slices stored in 0.25% O<sub>2</sub> atmosphere.

Key Words: zucchini-squash, fresh-cut, controlled atmosphere, respiration, storage quality

## INTRODUCTION

STUDIES have shown that controlled atmosphere (CA) extends the shelf life of some fruits and vegetables (Kader, 1986). Similarly, CA or modified atmosphere (MA) has been beneficial for some fresh-cut products such as chopped lettuce (McDonald et al., 1990), shredded cabbage (Kaji et al., 1993), broccoli spears (Barth et al., 1993), and strawberries (Larsen et al., 1995). In film packaged fresh-cut products, atmosphere can become modified extensively, with O<sub>2</sub> depleted below the extinction point. This may occur when the holding temperature is too high and/or permeability of gases through the film is inadequate. Lack of O<sub>2</sub> would result in anaerobiosis followed by injury to tissue. Information is lacking on the minimum O<sub>2</sub> level that fresh-cut products tolerate without undesirable effects and the quality changes that occur in very low O<sub>2</sub> atmospheres.

With whole zucchini squash fruit, a 1% O<sub>2</sub> atmosphere at 2.5°C (Wang and Ji, 1989) or 5% CO<sub>2</sub> at 5°C (Mencarelli, 1987) reduced chilling injury and subsequent decay. A 2% O<sub>2</sub> at 10°C was reported to minimize development of off flavor, which was more pronounced in cooked than in raw fruit (Mencarelli et al., 1983). Slices of zucchini squash may tolerate O<sub>2</sub> atmosphere of 1% because they do not have a continuous barrier to gases. The distance of gas diffusion is less in slices than whole fruit, which would result in a smaller gradient of O<sub>2</sub> (Brecht, 1980; Weichmann, 1986). Equally important is the determination of the minimum O<sub>2</sub> level that zucchini squash slices could tolerate in MA of film wrapped packages held at 5° to 10°C.

We previously reported that calcium dip treatment of zucchini squash slices reduced chilling injury at 0°C and retained quality at 10°C, where deterioration may occur readily (Izumi and Watada, 1995). Our objective was to determine the response of zucchini squash slices to O<sub>2</sub> levels <1% at 10°C and the effect of low O<sub>2</sub> on quality of calcium treated slices.

## MATERIALS & METHODS

ZUCCHINI SQUASH (*Cucurbita pepo* L. cv. Elite) were obtained from a farm in Clinton, MD, washed with water and trimmed

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at the stem and blossom ends. Trimmed fruit were sliced (5 mm thick, ca 5g each) transversely with a food processor (Model DLC-10, Cuisinarts, East Windsor, NJ).

## Response to low O<sub>2</sub> atmosphere storage

Samples (100g) of slices (3–4 cm diam, 5 mm thick) were placed in a 2L glass jar containing 100 mL distilled water in the bottom to maintain relative humidity >90%. Three replicated samples were stored at 10°C under a continuous humidified stream (15 mL/min) of air or low O<sub>2</sub> (0.25, 0.5 and 1%), with the balance being N<sub>2</sub>. Oxygen and CO<sub>2</sub> contents of inlet and outlet streams of each jar were monitored every 8 hrs with an O<sub>2</sub> and CO<sub>2</sub> analyzer (Model S-3A/I and Model CD-3A; Ametek, Pittsburgh, PA) and the average of 3 measurements/day was used for data analysis. A 5 mL aliquot of gas was taken daily and ethylene content was measured with a gas chromatograph (model AGC-211; Carle, Tulsa, OK) equipped with a flame ionization detector.

After 8 days storage, evaluations of browning/decay and off-odor were made for each sample. Data on brown discoloration and subsequent decay were combined in determining percentage of deteriorated slices. The severity of undesirable condition was reported as severity index, because tissues began to decay shortly after development of brown discoloration. Percentage of browning/decay was evaluated on a rating scale (R) of 0 = none, 1 = slight, 2 = moderate, 3 = severe, and 4 = extreme. The severity index (SI) is the weighted average:

$$SI = \frac{\sum N_i R_i}{N_T} \times 100$$

where N<sub>i</sub> is the number of slices with ranking R<sub>i</sub>, and N<sub>T</sub> is the total number of slices × the four scale ratings. This gives a score of severity on a scale ranging from 0 to 100 (Makita, 1985). Odor of slices was rated on a scale of 0 = normal to 4 = severely objectionable.

## Calcium treatment and low O<sub>2</sub> storage

A 500g sample of slices (3–4 cm diam and 5 mm thick) was dipped in 2L 0.5% CaCl<sub>2</sub> solution or distilled water for 2 min at room temperature (≈23°C) and centrifuged for 10 sec at 590 rpm to remove surface solution, as described earlier (Izumi and Watada, 1995). A 100g sample of slices was placed in a 2L glass jar containing 100 mL distilled water in the bottom. Three replicated samples were stored at 10°C under a continuous humidified stream (15 mL/min) of air or 0.25% O<sub>2</sub>. Oxygen, CO<sub>2</sub>, and ethylene levels of each sample were monitored as described.

An aliquot from each lot was removed periodically for determination of percentage and SI of browning/decay, score of off-odor, weight, L-ascorbic acid content, texture, total microbial count, pH, and color.

L-ascorbic acid content was determined using HPLC equipped with a PLRP-S 100A column (25 cm × 4.66 mm, 5 μm) (Polymer Laboratories, Co.) and electrochemical detector (Model 400; EG&G, Princeton, NJ) as described (Izumi and Watada, 1995). Texture, based on the force required to shear a 40g sample, and total microbial count on the surface of 10g tissues were determined as described (Izumi and Watada, 1994). The surface pH on both sides of a slice was measured with a pH meter (Model B-113; Horiba, Japan). The surface color of five slices for each sample was measured with a chroma meter (Model CR-300; Minolta, Japan) and results were expressed as L\* and hue angle (tan<sup>-1</sup>b\*/a\*) values (Izumi and Watada, 1995).

## Statistical analysis

Data were subjected to analysis of variance and Duncan's multiple range test for tabulated data, and the standard error of individual means was presented.

## RESULTS &amp; DISCUSSION

CARBON DIOXIDE production and O<sub>2</sub> consumption were lower with zucchini slices held in low O<sub>2</sub> than in air, and the rates of slices under different low O<sub>2</sub> atmospheres were similar (Fig. 1). With whole zucchini squash fruit, CO<sub>2</sub> production was reduced as O<sub>2</sub> level was reduced from that in air to 1% at 5° or 10°C (Mencarelli et al., 1983). The rates were lower as O<sub>2</sub> was reduced from 6 to 1%. The respiratory quotient (RQ) of slices held in 0.25% O<sub>2</sub> was similar to those at 0.5 or 1% O<sub>2</sub>, and at times was slightly higher than those held in air (data not shown). This indicated that sufficient O<sub>2</sub> was available in the low O<sub>2</sub> atmosphere for aerobic respiration to continue without initiating the anaerobic pathway.

Ethylene production was not detectable from slices in 0.25 and 0.5% O<sub>2</sub> and was minimal in 1.0% O<sub>2</sub>, thus its production was suspected to be inhibited by low O<sub>2</sub>. A similar type inhibition has been reported for whole fruit (Mencarelli et al., 1983). Apparently low O<sub>2</sub> reduces ethylene production by inhibiting the conversion of ACC to ethylene (Kader, 1986).

The zucchini squash slices developed brown discoloration and subsequently decayed during storage. This was probably due to natural deterioration. Percentage and severity of browning/decay were less under low O<sub>2</sub> than in air, and no differences were observed among various low O<sub>2</sub> atmospheres (Table 1). Low O<sub>2</sub> or high CO<sub>2</sub> atmospheres have been shown to reduce browning via inhibition of phenolics production and polyphenol oxidase activity with lettuce (Siriphanich and Kader, 1985) and shiitake mushroom (Minamide et al., 1980). Thus with zucchini slices, low O<sub>2</sub> may have inhibited some steps in the pathway of phenolic metabolism responsible for browning. CA conditions have been shown to suppress postharvest decay of fruits and vegetables by suppression of pathogenic growth of bacteria and fungi (El Goorani and Sommer, 1981). Low O<sub>2</sub> may have suppressed pathogen growth on the zucchini slices, which resulted in less decay. Off-odor was not induced by either low O<sub>2</sub> or air storage.

As noted with water treated slices, CO<sub>2</sub> production and O<sub>2</sub> consumption of calcium treated zucchini slices were lower when held in 0.25% O<sub>2</sub> than in air atmosphere, but no differences occurred between calcium and water treated slices (Fig. 2). The sharp increase in CO<sub>2</sub> production and O<sub>2</sub> consumption noted on day 7 by slices held in air coincided with the sharp increase in metabolic activity probably was due to deterioration and decay of tissue. By day 8, the respiration rate was about threefold greater than controls, which had minimal decay, suggesting decay had a notable effect on elevated metabolic rates. Increased metabolic rate or decay was not noted with calcium or water treated slices held in 0.25% O<sub>2</sub>. The RQ was higher in slices held in 0.25% O<sub>2</sub> than in air (Fig. 2) which may have resulted from increased glycolysis (Kader, 1986).

A 0.25% O<sub>2</sub> atmosphere seemed to inhibit the rate of ethylene production (Fig. 2). Treatment with calcium had no additive effect on the rates of respiration and ethylene production of slices held in 0.25% O<sub>2</sub>.

The percentage and severity of browning/decay in slices were suppressed by 0.25% O<sub>2</sub> regardless of calcium treatment (Table 2). Suppression by the 0.25% O<sub>2</sub> atmosphere may have been at the maximum, since the beneficial effect of calcium noted in a previous study (Izumi and Watada, 1995) was not noted when combined with the low O<sub>2</sub> treatment. Browning/decay in air samples differed widely between the two experiments (Table 1 and 2), which may have related to the difference in maturity of fruit (Mencarelli et al., 1983). Nevertheless, the low O<sub>2</sub> atmosphere was helpful in reducing the amount of browning/decay. Thus in commercial film-packed fresh-cut zucchinis, the reduced O<sub>2</sub> level caused by MA would be desirable in retarding development of browning/decay. Slices held in air developed off-odor on day 10 due to decay, while those slices held in 0.25% O<sub>2</sub> produced only a slight off-odor.

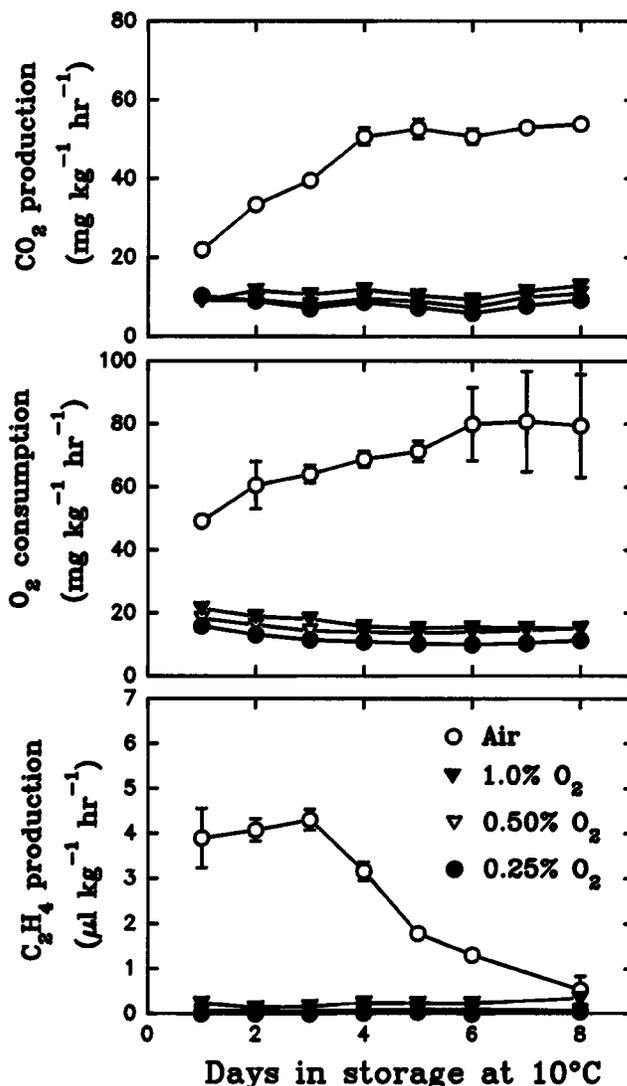


Fig. 1—Rates of gas changes in zucchini squash slices during storage at 10°C under air or low O<sub>2</sub> atmospheres. Vertical lines represent SE. SE bars were not shown when masked by the graph symbol.

Table 1—Some changes in zucchini squash slices after 8 days storage under air or low O<sub>2</sub> atmospheres at 10°C

Treatment	% of slices with browning/decay <sup>c</sup>	Severity index of browning/decay <sup>d</sup>	Odor <sup>e</sup>
Air	21.3 <sup>a</sup>	5.4 <sup>a</sup>	0 <sup>a</sup>
1%O <sub>2</sub>	3.6 <sup>b</sup>	0.9 <sup>b</sup>	0 <sup>a</sup>
0.5%O <sub>2</sub>	1.7 <sup>b</sup>	0.4 <sup>b</sup>	0.3 <sup>a</sup>
0.25%O <sub>2</sub>	1.8 <sup>b</sup>	0.4 <sup>b</sup>	0 <sup>a</sup>

<sup>ab</sup> Means with different superscripts in the same column are significantly different ( $p < 0.01$ ).

<sup>c</sup>  $\frac{\text{Number of browned and decayed slices}}{\text{Number of observed slices}} \times 100$ .

<sup>d</sup>  $\frac{(\text{Number of slight slices} \times 1) + (\text{Moderate slices} \times 2) + (\text{Severe slices} \times 3) + (\text{Extreme slices} \times 4)}{\text{Number of observed slices} \times 4} \times 100$ .

<sup>e</sup> Rated on a scale of 0 to 4, with 0 = normal and 4 = severely objectionable.

Total weight loss of both calcium treated and nontreated slices in 0.25% O<sub>2</sub> was reduced by 40% when compared to those in air (data not shown). Large differences in weight loss between samples in air and 0.25% O<sub>2</sub> atmospheres probably were due to differences in carbon loss in respiration (Fig. 2) and moisture loss. Slices held in air probably lost more water than those in low O<sub>2</sub> because of a difference in histology of tissues. Those in air were in a more advanced stage of senescence as noted by the deteriorated condition, which probably made them more susceptible to water loss.

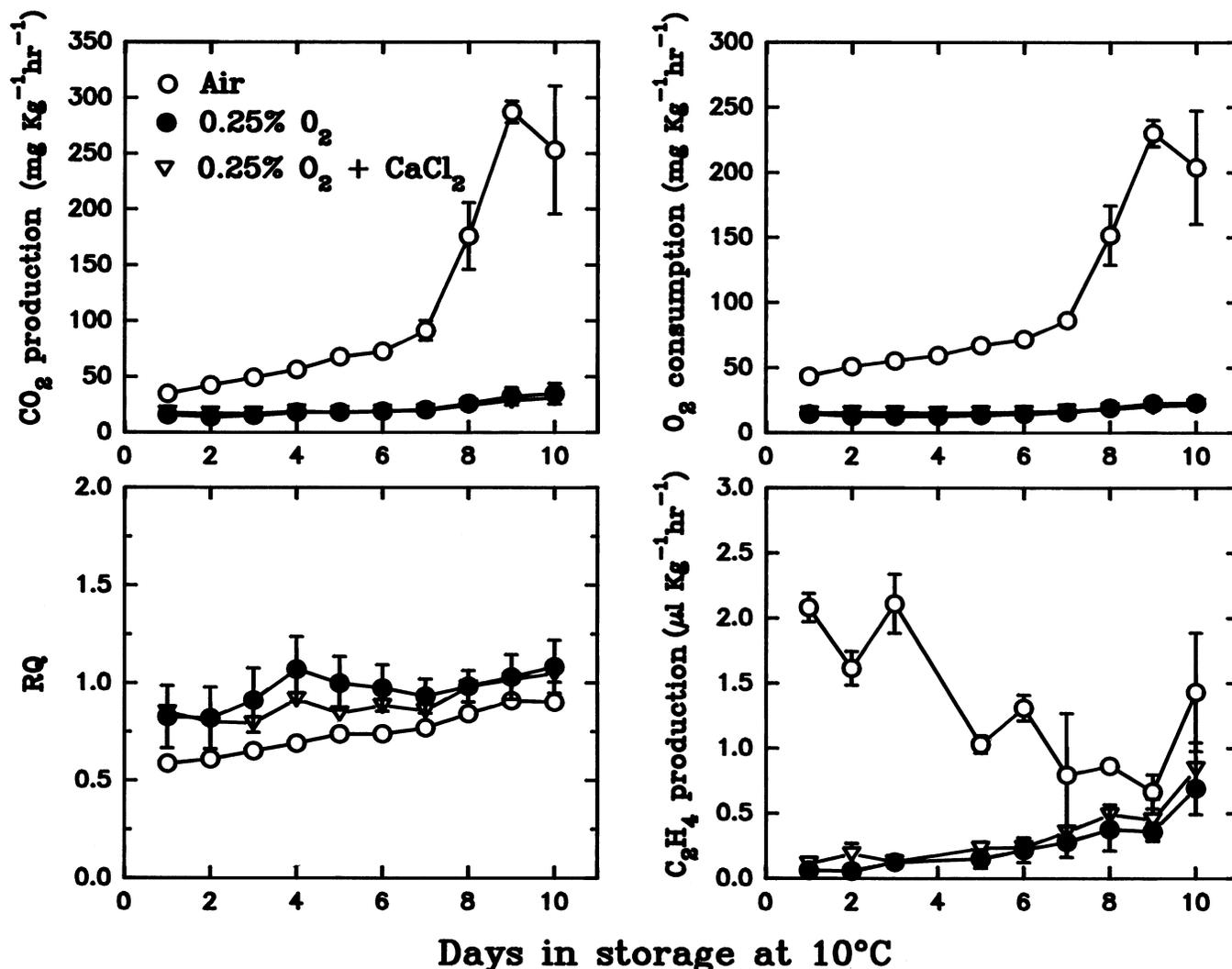


Fig. 2—Rates of gas changes and respiratory quotients (RO) of zucchini squash slices treated with CaCl<sub>2</sub> or water and stored in air or 0.25% O<sub>2</sub> at 10°C. Vertical lines represent SE. SE bars were not shown when masked by the graph symbol.

Table 2—Some changes in zucchini squash slices treated with CaCl<sub>2</sub> or water and stored in air or 0.25% O<sub>2</sub> at 10°C

Treatment	% of slices with browning/decay <sup>c</sup>			Severity index of browning/decay <sup>d</sup>			Odor <sup>e</sup>		
	3 <sup>f</sup>	7	10	3	7	10	3	7	10
Air	0 <sup>a</sup>	89.0 <sup>a</sup>	100.0 <sup>a</sup>	0 <sup>a</sup>	29.9 <sup>a</sup>	100.0 <sup>a</sup>	0 <sup>a</sup>	0 <sup>a</sup>	4 <sup>a</sup>
0.25%O <sub>2</sub>	0 <sup>a</sup>	10.7 <sup>b</sup>	55.6 <sup>b</sup>	0 <sup>a</sup>	2.7 <sup>b</sup>	23.7 <sup>b</sup>	0 <sup>a</sup>	0.3 <sup>a</sup>	1.3 <sup>b</sup>
0.25%O <sub>2</sub> +CaCl <sub>2</sub>	0 <sup>a</sup>	4.2 <sup>b</sup>	53.2 <sup>b</sup>	0 <sup>a</sup>	1.1 <sup>b</sup>	21.7 <sup>b</sup>	0 <sup>a</sup>	0.3 <sup>a</sup>	1.3 <sup>b</sup>

<sup>ab</sup> Means with different superscripts in the same column are significantly different ( $p < 0.01$ ).

<sup>c</sup>  $\frac{\text{Number of browned and decayed slices}}{\text{Number of observed slices}} \times 100$ .

<sup>d</sup>  $\frac{(\text{Number of slight slices} \times 1) + (\text{Moderate slices} \times 2) + (\text{Severe slices} \times 3) + (\text{Extreme slices} \times 4)}{\text{Number of observed slices} \times 4} \times 100$ .

<sup>e</sup> Rated on a scale of 0 to 4, with 0 = normal and 4 = severely objectionable.

<sup>f</sup> Days in storage.

Shear force of slices in 0.25% O<sub>2</sub> remained unchanged during storage, whereas those in air decreased after day 3 with development of browning/decay (data not shown). The calcium effect of maintaining the shear force index (Izumi and Watada, 1995) seemed to be masked by the beneficial effect of low O<sub>2</sub>. The influence of CA storage on texture of vegetables varies with the commodity (Weichmann, 1986) and the mechanism of CA effects is not fully understood (Kader, 1986). In our study, low O<sub>2</sub> atmosphere may have maintained the textural quality of zucchini slices by suppressing browning/decay and weight loss that would occur with senescence.

L-ascorbic acid content of slices in air decreased to about 15% of the initial value by day 10, while those in 0.25% O<sub>2</sub>

changed minimally (Fig. 3A). Low O<sub>2</sub> atmosphere has been shown to result in L-ascorbic acid retention in other vegetables (Weichmann, 1986; Klein, 1987) and a decrease in oxidation of L-ascorbic acid to dehydroascorbic acid (Weichmann, 1986). In our zucchini slices, better retention of L-ascorbic acid in 0.25% O<sub>2</sub> samples may have had an effect on reduction of browning (Table 2), because of the antioxidant property of L-ascorbic acid (Liao and Seib, 1987).

The total microbial count increased during storage and the count was higher for samples in air than in 0.25% O<sub>2</sub> on the last day (Fig. 3B). The lower count in the 0.25% O<sub>2</sub> samples may be a direct effect of O<sub>2</sub> below 1% having an inhibitory effect on growth and activities of pathogens and/or an indirect

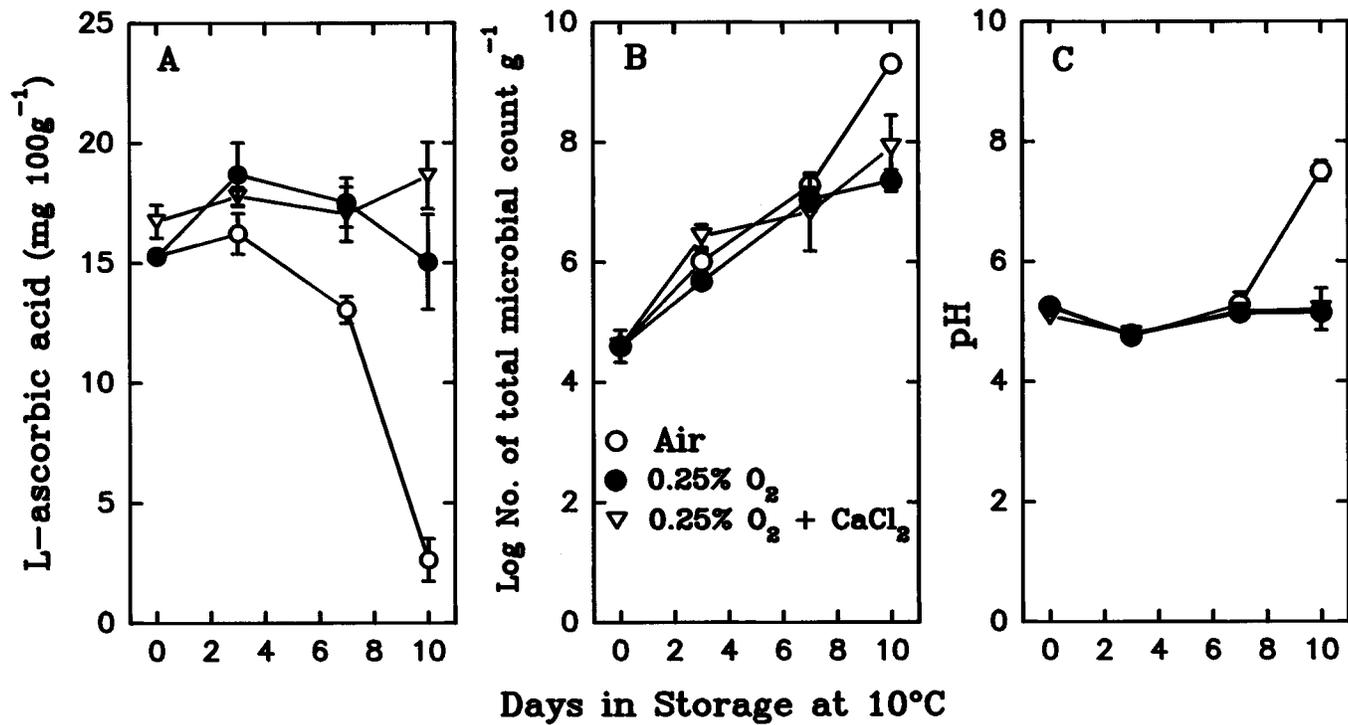


Fig. 3—Changes in L-ascorbic acid, microbial counts, and surface pH of zucchini squash slices treated with CaCl<sub>2</sub> or water and stored in air or 0.25% O<sub>2</sub> at 10°C. Vertical lines represent SE. SE bars were not shown when masked by the graph symbol.

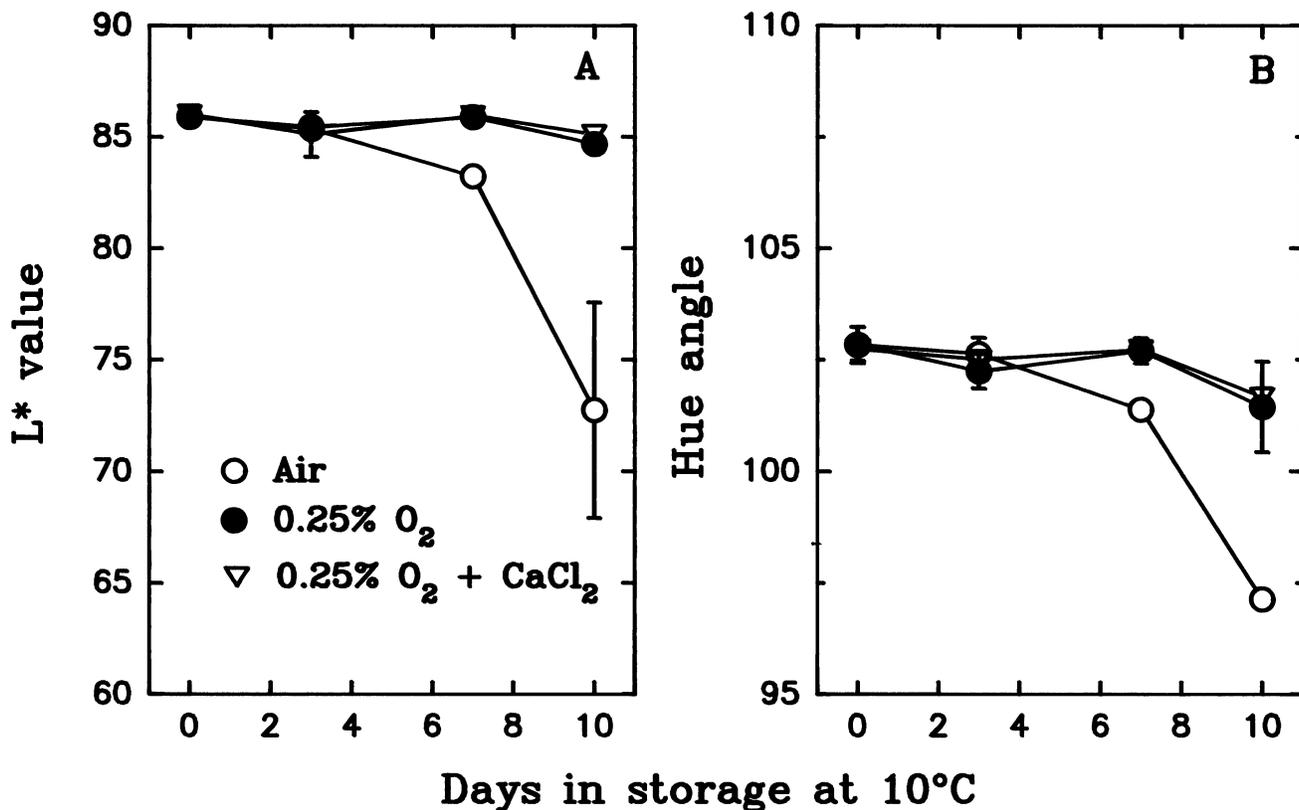


Fig. 4—Some color values of zucchini squash slices treated with CaCl<sub>2</sub> or water and stored in air or 0.25% O<sub>2</sub> at 10°C. Vertical lines represent SE. SE bars were not shown when masked by the graph symbol.

effect with the host commodity being less susceptible to pathogen growth. Ripening and senescence of fruit in low O<sub>2</sub> atmosphere are delayed relative to those in air, thus samples in low O<sub>2</sub> would not be as susceptible to infection as those in air (El-Goorani and Sommer, 1981).

The pH of slices held in air increased sharply on the last day which may have been associated with microbial growth (Fig. 3C). Microbial spoilage of CA packaged foods may be caused by several factors, which include pH, atmosphere, temperature, and water activity (Labuza et al., 1992).

Surface lightness, expressed as  $L^*$  values, was retained by the 0.25%  $O_2$  sample, whereas it decreased in the air sample after day 3 (Fig. 4A). Hue angle values of air samples also decreased due to the decrease in  $L^*$  values (Fig. 4B), which indicated that the green color was lost and yellow and shades of red were embodied in the mixture (McGuire, 1992). These brightness and color differences indicated that brownish discoloration was minimized by low  $O_2$  (Table 2). Treatment with calcium did not affect the quality attributes of zucchini squash slices held in low  $O_2$  atmosphere.

## CONCLUSIONS

Low  $O_2$  of 0.25 to 1% reduced respiration and ethylene production rates and retained visual quality of zucchini squash slices held at 10°C. A 0.25%  $O_2$  atmosphere had a beneficial effect on attributes including weight loss, development of browning/decay, shear force, L-ascorbic acid content, microbial count, pH, and color, and probably masked beneficial effects of calcium dip. Zucchini squash slices were not deleteriously affected by 0.25%  $O_2$  atmosphere, so  $O_2$  levels in the film packaged slices could be allowed to reach this level.

## REFERENCES

Barth, M.M., Kerbel, E.L., Perry, A.K., and Schmidt, S.J. 1993. Modified atmosphere packaging affects ascorbic acid, enzyme activity and market quality of broccoli. *J. Food Sci.* 58: 140–143.  
 Brecht, P.E. 1980. Use of controlled atmospheres to retard deterioration of produce. *Food Technol.* 34(3): 45–50.  
 El-Goorani, M.A. and Sommer, N.F. 1981. Effects of modified atmospheres on postharvest pathogens of fruits and vegetables. *Hort. Rev.* 3: 412–461.  
 Izumi, H. and Watada, A.E. 1994. Calcium treatments affect storage quality of shredded carrots. *J. Food Sci.* 59: 106–109.  
 Izumi, H. and Watada, A.E. 1995. Calcium treatment to maintain quality of zucchini squash slices. *J. Food Sci.* 60: 789–793.  
 Kader, A.A. 1986. Biochemical and physiological basis for effects of controlled and modified atmospheres on fruits and vegetables. *Food Technol.* 40(5): 99–104.

Kaji, H., Ueno, M., and Osajima, Y. 1993. Storage of shredded cabbage under a dynamically controlled atmosphere of high oxygen and high carbon dioxide. *Biosci. Biotechnol. Biochem.* 57: 1049–1052.  
 Klein, B.P. 1987. Nutritional consequences of minimal processing of fruits and vegetables. *J. Food Qual.* 10: 179–193.  
 Labuza, T.P., Fu, B., and Taoukis, P.S. 1992. Prediction for shelf life and safety of minimally processed CAP/MAP chilled foods: A review. *J. Food Prot.* 55: 741–750.  
 Larsen, M. and Watkins, C.B. 1995. Firmness and concentrations of acetaldehyde, ethyl acetate and ethanol in strawberries stored in controlled and modified atmospheres. *Postharvest Biol. & Technol.* 5: 39–50.  
 Liao, M. and Seib, P.A. 1987. Selected reactions of L-ascorbic acid related to foods. *Food Technol.* 41(11): 104–107, 111.  
 Makita, Y. 1985. Histochemical observation on the pitting rind of some citrus varieties and effects of prestorage hot-water treatments on the prevention of rind disorder. *Bull. Shizuoka Citrus Exp. Sta.* 21: 45–47.  
 McDonald, R.E., Risse, L.A., and Barmore C.R. 1990. Bagging chopped lettuce in selected permeability films. *HortScience* 25: 671–673.  
 McGuire, R.G. 1992. Reporting of objective color measurements. *HortScience* 27: 1254–1255.  
 Mencarelli, F. 1987. Effect of high  $CO_2$  atmospheres on stored zucchini squash. *J. Amer. Hort. Sci.* 112: 985–988.  
 Mencarelli, F., Lipton, W.J., and Peterson, S.J. 1983. Responses of 'Zucchini' squash to storage in low- $O_2$  atmospheres at chilling and nonchilling temperatures. *J. Amer. Soc. Hort. Sci.* 108: 884–890.  
 Minamide, T., Nishikawa, T., and Ogata, K. 1980. Influences of  $CO_2$  and  $O_2$  on the keeping freshness of shii-take (*Lentinus edodes* (Berk) Sing.) after harvest. *J. Japan. Soc. Food Sci. Technol.* 27: 505–510.  
 Siriphanich, J. and Kader, A.A. 1985. Effects of  $CO_2$  on total phenolics, phenylalanine ammonia lyase, and polyphenol oxidase in lettuce tissue. *J. Amer. Soc. Hort. Sci.* 110: 249–253.  
 Wang, C.Y. and Ji, Z.L. 1989. Effect of low-oxygen storage on chilling injury and polyamines in zucchini squash. *Sci. Hort.* 39: 1–7.  
 Weichmann, J. 1986. The effect of controlled-atmosphere storage on the sensory and nutritional quality of fruits and vegetables. *Hort. Rev.* 8: 101–127.  
 Wills, R.B.H., Wimalasiri, P., and Scott, K.J. 1979. Short pre-storage exposures to high carbon dioxide or low oxygen atmospheres for the storage of some vegetables. *HortScience* 14: 528–530.  
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Dawson, D.M., Watkins, C.B., and Melton, L.D. 1993. Calcium uptake and efflux, ion leakage, internal air space and cation exchange capacity in relation to mealiness in nectarine tissue. *Postharvest Biol. Technol.* 3: 131–141.  
 De Haan, I. 1957. Pectin conversion in peaches during cold storage. *South African Ind. Chem.* 11: 26–34.  
 Downs, C. and Brady, C.J. 1990. Two form of exopolygalacturonase increased as peach fruit ripen. *Plant Cell Environ.* 13: 523–530.  
 Downs, C.G., Brady, C.J., and Gooley, A. 1992. Exopolygalacturonase protein accumulates late in peach fruit ripening. *Physiol. Plant.* 85: 133–140.  
 Fry, C.F. 1986. Cross-linking of matrix polymers in the growing cell walls of angiosperms. *Annu. Rev. Plant Physiol.* 37: 165–186.  
 Huber, D.J. 1983. The role of cell wall hydrolase in fruit softening. *Hort. Reviews* 5: 169–219.  
 Jarvis, M.C. 1984. Structures and properties of pectin gels in plant cell walls. *Plant Cell Environ.* 7: 153–164.  
 Kramer, G.F., Wang, C.Y., and Conway, W.S. 1989. Correlation of reduced softening and increased polyamine levels during low-oxygen storage of 'McIntosh' apples. *J. Am. Soc. Hort. Sci.* 114: 942–946.  
 Lill, R.E., O'Donoghue, E.M., and King, G.A. 1989. Postharvest physiology of peaches and nectarines. *Hort. Rev.* 11: 413–452.  
 McDonald, R.E. and Kushad, M.M. 1986. Accumulation of putrescine during chilling injury of fruits. *Plant Physiol.* 82: 324–326.  
 Nelson, N. 1944. A photometric adaptation of the Somogyi method for the determination of glucose. *J. Biol. Chem.* 153: 375–380.

Pozsár-Hajnal, K. and Polacsek-Rácz, M. 1975. Determination of pectinmethylesterase, polygalacturonase and pectin substances in some fruits and vegetables. *Acta Alimentaria* 4: 271–289.  
 Pressey, R. and Avants, J.K. 1973. Separation and characterization of endopolygalacturonase and exopolygalacturonase from peaches. *Plant Physiol.* 52: 252–256.  
 Pressey, R. and Avants, J.K. 1978. Difference in polygalacturonase composition of clingstone and freestone peaches. *J. Food Sci.* 43: 1415–1417, 1423.  
 Strand, L.L., Rehtoris, C., and Mussell, H. 1976. Polygalacturonase releases cell-wall bound proteins. *Plant Physiol.* 58: 722–725.  
 von Mollendorff, L.J. and de Villiers, O.T. 1988. Role of pectolytic enzymes in the development of woolliness in peaches. *J. Hort. Sci.* 63: 53–58.  
 Wheatley, D.N. 1984. Intracellular protein degradation: Bases for a self-regulating mechanism for the proteolysis of endogenous proteins. *J. Theor. Biol.* 107: 127–149.  
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