Lithium metaborate fusion for silicon, calcium, magnesium, and potassium analysis of wild rice

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Abstract

A rapid batch method was developed for the analysis of Si, Ca, Mg, and K in a large number of plant tissue samples by fusion with lithium metaborate (LiBO₃) in graphite crucibles with the use of a molybdenum blue spectrophotometric analysis of silicon and FAAS for Ca, Mg, and K. Our method was tested for whole plant analysis of mature wild rice (*Zizania palustris* L.). Analysis of Si in plant tissue with LiBO₃ in graphite crucibles is reliable and fast. Thirty-six samples can be ashed overnight, fused in one hour the next day, and analyzed for Si within approximately two hours.

Introduction

Analysis of silicon in plant tissue may be accomplished using dissolution techniques and solution analysis or gravimetrically after ashing and extraction of the soluble components of the ash. The former requires the complete dissolution of biogenic opal and possible SiO2/organic complexes (Novozamsky, 1984). This can be accomplished by digestion with mixtures of HNO3-HClO₄-HF (Wikoff, 1986), extraction with HCl-HF without wet or dry ashing of organic material (Novozamsky, 1984), ashing with concentrated H2SO4 in platinum crucibles and fusion with Na₂CO₃ (Volk and Weintraub, 1958), and ashing overnight at 550 °C in Ni crucibles followed by fusion with NaOH (Fox et al., 1969).

Most fusion techniques require heating individual samples over a flame to a dull redness in Pt crucibles for approximately 10 minutes to ensure complete reaction of the silica with the molten salt. The fused material is then dissolved in acid. Silicon can be determined by a silicomolybdenum blue spectrophotometric method,

flame atomic absorption spectrometry (FAAS), or, by inductively coupled plasma atomic emission spectrometry (ICP-AES). Fusion in crucibles, however, is time consuming, especially for a large number of samples, and Pt crucibles are expensive.

Digestion (wet ashing) requires destruction of plant organic matter with an oxidizing acid and dissolution of silica with HF followed by FAAS analysis or ICP-AES using an HF resistant nebulizer assembly. The commonly used silicomolybdenum blue method is not suitable for the determination of Si following digestion with HF because F strongly complexes silica. This procedure is flexible enough to accommodate processing of large numbers of samples in a relatively short period of time. However, a typical complaint about this method is the build-up of carbon on the FAAS burner head (Hallmark et al., 1982). This method also requires the use of HF and special procedures to prevent loss of volatile SiF₁.

A rapid gravimetric procedure for determination of Si in plant tissue requiring the oxidation of plant tissue was developed by Elliott et

al. (1988). With this procedure, finely ground dried plant tissue is placed in fritted-glass Gooche crucibles, washed with 2 M HCl, H₂O, and acetone, and dried at 80 °C. The sample is ashed at 550 °C, cooled, washed as above, and dried. The remaining mass after all treatment is considered to be silica. The procedure is rapid, but requires a separate digestion for Ca, Mg, K, and other plant nutrients.

Determination of Ca, Mg, and K in plant tissue is often accomplished by digestion with mixtures of HNO₃-HClO₄ or HNO₃-HClO₄-H₂SO₄ (Wikoff, 1986), digestion with H₂SO₄-H₂O₂ (Lowther, 1980), or by ashing at 550 °C followed by addition of HCl or HNO₃ (Munter, 1984). Silicon cannot be directly determined by any of these methods.

Our objective was to develop a batch method for the rapid analysis of Si, Ca, Mg, and K in a large number of mature wild rice (Zizania palustris L.) samples by fusion with lithium metaborate (LiBO3) in graphite crucibles with the use of a molybdenum blue spectrophotometric analysis of silicon and FAAS for Ca, Mg, and K. Our method is an adaptation of techniques used for the analysis of silicate minerals (Feldman, 1983; Khalid et al., 1978; Suhr and Ingamells) and estuarian sediment (Cantillo et al., 1984). Their procedure for analysis of silicate minerals was as to carefully mix 100 mg of material with 500 mg of lithium metaborate in tared graphite crucibles (Suhr and Ingamells, 1966). The sample was heated to 950 °C for 10 to 15 minutes. The molten flux was transferred to $50\,\text{mL}$ of 3%HNO3 in a teflon container and stirred until solution was complete. Cantillo et al. (1984) followed essentially the same procedure with estuarian sediment. They found that incomplete fusion may result if sediment comes in direct contact with the graphite crucible.

Materials and methods

Wild rice (Zizania palustris L.) whole plant tissue samples were obtained from a CaSiO₄ fertility experiment in a peat soil. Four treatments of CaSiO₄ were applied, 0.0 (controls), 2250, 6750, and 13500 kg ha $^{-1}$ with four replications. Three treatments of CaCO3 (agricultural

limestone) were applied as Ca and pH controls at the same rates as CaSiO₄ with four replications.

At physiological maturity, plants from 1.0 m² area subplots, 3 per plot, were harvested and dried at 60 °C. Whole plant tissue samples were prepared for analysis by milling samples to pass a 1-mm sieve.

Si analysis: Lithium metaborate fusion

Graphite crucibles (SPEX Industries), 10 mL capacity, were tared and 300 mg of LiBO3 was added. One hundred milligrams of plant tissue was added, taking care that the sample was not in contact with the wall of the crucible. Eighteen to twenty crucibles were placed on flat stainless steel trays (14 gage) 14 cm by 28 cm. The edges of the trays were bent upward to provide a border one cm high to prevent the crucibles from sliding off the trays.

The trays were placed in a muffle furnace and heated slowly to 485 °C over a period of five hours to ash the plant tissue. Heating was continued for 24 hours at 485 °C. The trays were removed from the oven and the oven temperature was increased to 950 °C. During this time the plant ash was carefully mixed into the surface of the LiBO3 with a small clean spatula. With large long-handled (40 to 50 cm) tongs and leather gloves one tray was placed in the furnace for approximately 15 minutes or until the temperature returned to 950 °C. The tray was removed and placed on a heat resistant slab to cool. The fused LiBO3 and plant ash formed a small solid spherical bead which was dropped in 50 mL or 100 mL of 2 M HCl in a high density polyethylene plastic bottle prepared by washing with 0.1 M HCl. Occasionally, small pieces of fused LiBO3 were not consolidated with the spherical bead. However, they were easily transferred to the 2M HCl. Pouring molten LiBO₃, as suggested by Cantillo et al. (1984) was difficult, dangerous, and not necessarily. Larger volumes of acid can be used if the sample is expected to contain larger amounts of silica.

The bottles were capped and placed on a shaker for two hours. Dissolution appeared to be complete in 30 to 40 minutes. Appropriate dilutions were made and the silicon content was determined by the molybdenum-blue method (Hallmark et al., 1982). Precision for the lithium

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metaborate fusion method in graphite crucibles was tested in ten wild rice tissues samples analyzed in triplicate for Si over a six month period. Samples included husks, stems, leaves, and whole plant samples. Accuracy and precision of the fusion method was determined by fusion of three samples of amorphous silica ($\rm H_2O < 1\%$) which is chemically similar to opal phytoliths. Precision for the gravimetric method was determined by duplicate analysis of wild rice samples from the CaSiO₄ experiment.

Subsamples of the plant tissue were sent for Si analysis to Dr. George Snyder at the University of Florida Agriculture and Research Center at Belle Glade for comparison with our results. Si was determined gravimetrically according to Elliot et al. (1988).

Ca, Mg, and K analysis: Lithium metaborate fusion

Wild rice was obtained from a nitrogen response study conducted in a greenhouse with pea soil. Growth and development was highly variable among treatments. Two plants were harvested at maturity from each pot, dried, and milled. Lithium metaborate fusion was conducted on all samples. Diluted solutions were analyzed for Ca, and Mg by FAAS and for K by flame emission spectrometry with standard prepared on the same solution matrix as the Si samples.

For comparison with the standard dry asing procedure used in our laboratory, dried plant samples (500 mg) from the nitrogen response study were placed in quartz crucibles and ashed at 550 °C. The ash was treated with 2 M HCl and analyzed for Ca, Mg, and K by inductively coupled plasma emission spectrometry (ICP–AES) (Munter, 1984).

Statistical analysis was conducted using STATISTIX, a microcomputer program (NH Analytical Software, Roseville, MN). Confidence intervals and significance at the 95% level.

Results and discussion

The accuracy and precision for the determination of Si in amorphous silica, with a Si content of 46.7%, was 45.5% with a coefficient of variation of 4.7% (n = 3). The triplicate analysis of 10 wild rice samples with Si contents of 1.3 to 4.7% resulted in coefficients of variation ranging from 1.1 to 8.2% for analysis repeated over a period of several months. Improvement in precision for Si could probably be achieved with a combination of finer milling of plant tissue and ashing of 200 mg rather than 100 mg of sample.

A comparison of the lithium metaborte results for Si in wild rice tissue with the gravimetric method is shown in Figure 1 ($r^2 = 0.95$, SE = 0.22). The slope and intercept, with confidence

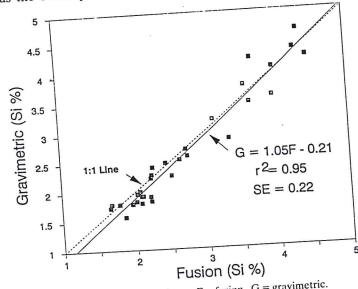


Fig. 1. Si in wild rice tissue. F = fusion, G = gravimetric.

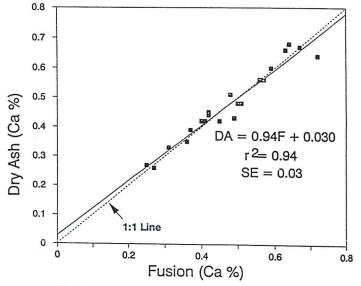


Fig. 2. Ca in wild rice tissue: dry ash vs. fusion.

intervals (Table 1),, indicate no significant difference between the two analytical techniques.

The plot in Figure 1 and the intercept value in Table 1, however, suggest a tendency for Si concentrations to be lower with the gravimetric method at low plant Si content. Perhaps the small amount of Si dissolved in the washing is significant for the low ash samples.

Results of Ca, Mg, and K analysis by dry ashing and ICP analysis vs. lithium metaborate fusion and FAAS are presented in Figures 2, 3, and 4, respectively, with the coefficient of simple

determination (r²) and standard error of mean (SE). Linear regression analysis indicates that both methods provide similar results for the elemental content of wild rice tissue. Slopes and intercepts are equal to 1.0 and 0.0, respectively, for Ca and Mg. The slope for the K plot is 1.0, the intercept confidence interval of the K plot is 0.029 to 0.195, suggesting that for very low K contents the dry ash procedure may give slightly higher values.

Analysis of Si in plant tissue with LiBO₃ in graphite crucibles is reliable and fast. Thirty-six

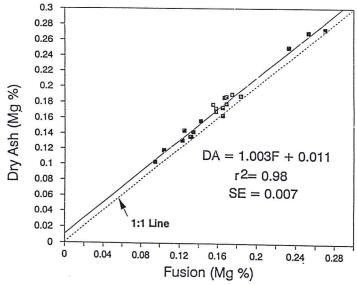


Fig. 3. Mg in wild rice tissue: dry ash vs. fusion.

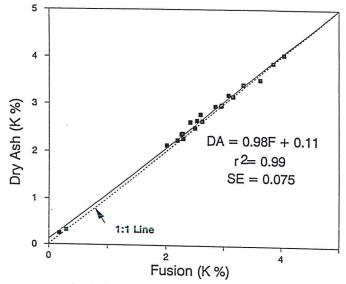


Fig. 4. K in wild rice tissue: dry ash vs. fusion.

Table 1. Linear regression results comparing dry ashing and ICP analysis vs. lithium metaborate fusion and FAAS analysis for Si, Mg, Ca, and K

Element	Regression analysis	Slope(±)	Intercept(±)
Si	G = 1.05 F - 0.21	0.093	0.27
Ca	DA = 0.9 F + 0.03	0.09	0.04
Mg	DA = 1.003 F + 0.011	0.057	0.042
K	DA = 0.98 F + 0.11	0.03	0.083

G = gravimetric analysis.

DA = dry ash.

F = samples prepared by LiBO₃ fusion.

samples can be ashed overnight, fused in one hour the next day, and analyzed for Si within approximately two hours. The technique can also be used for analysis of Ca, Mg, and K in plant tissue.

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